

# 1 **Jumping back and forth: anthropozoonotic and zoonotic** 2 **transmission of SARS-CoV-2 on mink farms**

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23

24 **One sentence summary**

25 SARS-CoV-2 transmission on mink farms.

26

27 **Abstract**

28 The zoonotic origin of the SARS-CoV-2 pandemic is still unknown. Animal experiments have  
29 shown that non-human primates, cats, ferrets, hamsters, rabbits and bats can be infected by  
30 SARS-CoV-2. In addition, SARS-CoV-2 RNA has been detected in felids, mink and dogs in the  
31 field. Here, we describe an in-depth investigation of outbreaks on 16 mink farms and humans  
32 living or working on these farms, using whole genome sequencing. We conclude that the virus  
33 was initially introduced from humans and has evolved, most likely reflecting widespread  
34 circulation among mink in the beginning of the infection period several weeks prior to  
35 detection. At the moment, despite enhanced biosecurity, early warning surveillance and  
36 immediate culling of infected farms, there is ongoing transmission between mink farms with  
37 three big transmission clusters with unknown modes of transmission. We also describe the  
38 first animal to human transmissions of SARS-CoV-2 in mink farms.

39

## 40 **Main text**

41 Late December 2019, SARS-CoV-2 was identified as the causative agent in a viral pneumonia  
42 outbreak, possibly related to a seafood and a live animal market in Wuhan, China (1). Since  
43 then, SARS-CoV-2 spread across the world and by August 31<sup>st</sup>, over 25,200,000 people had  
44 been infected with SARS-CoV-2 resulting in over 840,000 deaths (2). In the Netherlands, over  
45 72,000 infections have been confirmed, over 6,200 SARS-CoV-2 related deaths have been  
46 reported, and drastic measures have been put into place to prevent further spread of SARS-  
47 CoV-2 (3).

48 In view of the similarities with SARS-CoV-1, a zoonotic origin of the outbreak was  
49 suspected by the possible link with the Wuhan market where various animals were sold  
50 including fish, shellfish, poultry, wild birds and exotic animals. The finding of cases with onset  
51 of illness well before the period observed in the cluster, however, suggests the possibility of  
52 other sources (4). Although closely related coronaviruses in bats (5, 6) and pangolins (7, 8)  
53 have most sequence identity to SARS-CoV-2, the most likely diversion date of SARS-CoV-2  
54 from the most closely related bat sequence is estimated to date back to somewhere between  
55 1948-1982 (9). Therefore, the animal reservoir(s) of SARS-CoV-2 is (are) yet to be identified.

56 Experimental infections in dogs (10), cats (10, 11), ferrets (10, 12), hamsters (13, 14),  
57 rhesus macaques (15), tree shrew (16), cynomolgus macaques (17), grivets (18), common  
58 marmosets (19), rabbits (20), and fruit bats (21) have shown that these species are susceptible  
59 to SARS-CoV-2, and experimentally infected cats, tree shrews, hamsters and ferrets could  
60 transmit the virus. In contrast, experimental infection of pigs and several poultry species with  
61 SARS-CoV-2 proved to be unsuccessful (10, 21, 22). SARS-CoV-2 has also sporadically been  
62 identified in naturally infected animals. In the USA and in Hong Kong, SARS-CoV-2 RNA has  
63 been detected in dogs (23). In the Netherlands, France, Hong Kong, Belgium and the USA, cats

64 have tested positive by RT-PCR for SARS-CoV-2 (24–27). Furthermore, SARS-CoV-2 has been  
65 detected in four tigers and three lions in a zoo in New York (28). In Italy, the Netherlands and  
66 in Wuhan, antibodies to SARS-CoV-2 have been detected in cats (29–31). Recently, detected  
67 SARS-CoV-2 was detected in farmed mink (*Neovison vison*) that showed signs of respiratory  
68 disease and increased mortality (29, 32).

69         Thereafter, the Dutch national response system for zoonotic diseases was activated,  
70 and it was concluded that the public health risk of animal infection with SARS-CoV-2 was low,  
71 but that there was a need for increased awareness of possible involvement of animals in the  
72 COVID-19 epidemic. Therefore, from May 20<sup>th</sup> 2020 onwards, mink farmers, veterinarians and  
73 laboratories were obliged to report symptoms in mink (family *Mustelidae*) to the Netherlands  
74 Food and Consumer Product Safety Authority (NFCPSA) and an extensive surveillance system  
75 was set up (33).

76         Whole genome sequencing (WGS) can be used to monitor the emergence and spread  
77 of pathogens (34–37). As part of the surveillance effort in the Netherlands over 1,750 SARS-  
78 CoV-2 viruses have been sequenced to date from patients from different parts of the  
79 Netherlands (38). Here, we describe an in-depth investigation into the SARS-CoV-2 outbreak  
80 in mink farms and mink farm employees in the Netherlands, combining epidemiological  
81 information, surveillance data and WGS on the human-animal interface.

82

## 83 **Methods**

### 84 **Outbreak investigation**

85 Following initial detection of SARS-CoV-2 in mink on two farms on April 23<sup>rd</sup> and April 25<sup>th</sup>,  
86 respectively, as part of routine health monitoring done by the Royal GD Animal Health service  
87 and subsequent investigation by Wageningen Bioveterinary Research (WBVR), the national  
88 reference laboratory for notifiable animal diseases, a One Health outbreak investigation team  
89 was convened (39, 40). Subsequently, respiratory signs and increased mortality in mink was  
90 made notifiable by the Dutch Ministry of Agriculture, Nature and Food Quality and the farms  
91 were quarantined (no movements of animals and manure and visitor restrictions). On May 7<sup>th</sup>  
92 two other mink farms in the same region were confirmed to be infected. By the end of May  
93 the Dutch minister of Agriculture decided that all mink on SARS-CoV-2 infected farms had to  
94 be culled. Moreover, as the clinical manifestation of the infection was highly variable within  
95 and between farms, including asymptomatic infections, weekly testing of dead animals for  
96 SARS-CoV-2 infections became compulsory for all mink farms in the Netherlands. Moreover,  
97 a nation-wide transport ban of mink and mink manure, and a strict hygiene and visitor  
98 protocol was implemented. The first infected mink farms were culled from June 6<sup>th</sup> onwards.  
99 From the 10<sup>th</sup> infected farm (NB10) onwards, culling took place within 1-3 days after diagnosis.  
100 In this manuscript, the data up to June 26<sup>th</sup>, when a total of 16 mink farms in the Netherlands  
101 were found positive for SARS-CoV-2 infections, is presented.

102

### 103 **Veterinary and human contact tracing**

104 The Netherlands Food and Consumer Product Safety Authority (NVWA) traced animal related  
105 contacts with other mink farms. Backward and forward tracing of possible high-risk contacts  
106 was done in the framework of the standard epidemiological investigation by the NVWA (i.e.

107 focused on movement of vehicles, visitors such as veterinary practitioners, (temporary)  
108 workers, sharing of equipment between farms and transport and delivery of materials, such  
109 as feed, pelts, carcasses and manure). Persons with possible exposure from this investigation,  
110 as well as farm owners and resident farm workers were asked to report health complaints to  
111 the municipal health service for testing and – in the case of confirmed infections – for health  
112 advice and further contact tracing. Farm owners and workers on infected mink farms were  
113 informed of potential risks and were given advice on the importance and use of personal  
114 protective equipment and hygiene when handling animals (41). The contact structure on the  
115 farms was assessed through in-depth interviews, to identify additional persons with possible  
116 exposure to mink. In order to provide an enhanced set of reference genome sequences,  
117 anonymized samples from patients that had been diagnosed with COVID-19 in the area of the  
118 same four-digits postal codes as farms NB1-NB4 in March and April 2020 were retrieved from  
119 clinical laboratories in the region.

120

### 121 **SARS-CoV-2 diagnostics and sequencing**

122 The presence of viral RNA in mink samples was determined using a RT-PCR targeting the E  
123 gene as previously described (42). For the human samples, diagnostic RT-PCR was performed  
124 for the E and the RdRp gene (42). In addition, serology was performed, using the Wantai Ig  
125 total and IgM ELISA, following the manufacturer's instructions(43). For all samples with a Ct  
126 value <32, sequencing was performed using a SARS-CoV-2 specific multiplex PCR for Nanopore  
127 sequencing, as previously described (3). The libraries were generated using the native barcode  
128 kits from Nanopore (EXP-NBD104 and EXP-NBD114 and SQK-LSK109) and sequenced on a R9.4  
129 flow cell multiplexing 24 samples per sequence run. Flow cells were washed and reused until  
130 less than 800 pores were active. The resulting raw sequence data was demultiplexed using

131 Porechop (<https://github.com/rrwick/Porechop>). Primers were trimmed after which a  
132 reference-based alignment was performed. The consensus genome was extracted and  
133 positions with a coverage <30 were replaced with an “N” as described previously (44).  
134 Mutations in the genome compared to the GISAID sequence EPI\_ISL\_412973 were confirmed  
135 by manually checking the mapped reads and homopolymeric regions were manually checked  
136 and resolved by consulting reference genomes. The average SNP difference was determined  
137 using snp-dists (<https://github.com/tseemann/snp-dists>). All sequences generated in this  
138 study are available on GISAID.

139

#### 140 **Phylogenetic analysis**

141 All available near full-length Dutch SARS-CoV-2 genomes available on 1<sup>st</sup> of July were selected  
142 (n=1,775) and aligned with the sequences from this study using MUSCLE (45). Sequences with  
143 >10% “Ns” were excluded. The alignment was manually checked for discrepancies after which  
144 IQ-TREE (46) was used to perform a maximum likelihood phylogenetic analysis under the  
145 GTR+F+I +G4 model as best predicted model using the ultrafast bootstrap option with 1,000  
146 replicates. The phylogenetic trees were visualized in Figtree  
147 (<http://tree.bio.ed.ac.uk/software/figtree/>). For clarity reasons all bootstrap values below 80  
148 were removed. To look at potential relationships with migrant workers, also all Polish  
149 sequences from GISAID (47) were included in the alignment (**Supplementary table 1**).

150

#### 151 **Mapping specific mutation patterns on mink farms and in mink farm employees**

152 Amino acid coordinates are described in relation to the Genbank NC\_045512.2 reference  
153 genome. Open reading frames were extracted from the genome alignment using the genome  
154 annotation as supplied with the reference genome. A custom R script was used to distinguish



155 synonymous from non-synonymous mutations and non-synonymous mutations were  
156 visualized using a tile map from the ggplot2 package (48).

157

### 158 **Geographical overview of mink farms in the Netherlands and SARS-CoV-2 positive farms**

159 To protect confidentiality, SARS-CoV-2 positive mink farms were aggregated at municipality  
160 level. The datasets “*Landbouw; gewassen, dieren en grondgebruik naar gemeente*” and “*Wijk-  
161 en Buurkaart 2019*” from Statistics Netherlands (CBS) were used (49). Maps were created  
162 using R packages sp (50), raster (51) and rgdal (52) and ArcGIS 10.6 software by ESRI.

163

## 164 Results

165 SARS-CoV-2 was first diagnosed on two mink farms in the Netherlands on April 23<sup>rd</sup> (NB1) and  
166 April 25<sup>th</sup> (NB2), respectively. After the initial detection of SARS-CoV-2 on these farms an in-  
167 depth investigation was started to look for potential transmission routes and to perform an  
168 environmental and occupational risk assessment. Here, we describe the results of the  
169 outbreak investigation of the first 16 SARS-CoV-2 infected mink farms by combining SARS-CoV-  
170 2 diagnostics, WGS and in-depth interviews.

171

### 172 Screening of farm workers and contacts

173 Farm owners of the 16 SARS-CoV-2 positive mink farms were contacted by the municipal  
174 health services to conduct contact investigation and samples were taken for RT-PCR-based  
175 and serological SARS-CoV-2 diagnostics. In total, 97 individuals were tested by either  
176 serological assays and/or RT-PCR. In total, 43 out of 88 (49%) upper-respiratory tract samples  
177 tested positive by RT-PCR while 38 out of 75 (51%) serum samples tested positive for SARS-  
178 CoV-2 specific antibodies. In total, 66 of 97 (67%) of the persons tested had evidence for SARS-  
179 CoV-2 infection (**table 1**).

180

181 **Table 1.** Overview of human sampling on SARS-CoV-2 positive mink farms.

Farm:	First diagnosis in animals:	Date(s) of sampling employees and family members:	PCR positive (%)	Serology positive (%)	Employees and family members tested positive (PCR and/or serology)
NB1	24-04-2020	28-04-2020 – 11-05-2020	5/6 (83%)	5/5 (100%)	6/6 (100%)
NB2	25-04-2020	31-03-2020 – 30-04-2020	1/2 (50%)	8/8 (100%)	8/8 (100%)
NB3	07-05-2020	11-05-2020 – 26-05-2020	5/7 (71%)	0/6 (0%)*	5/7 (71%)
NB4	07-05-2020	08-05-2020	1/3 (33%)	2/2 (100%)	2/3 (66%)
NB5	31-05-2020	01-06-2020	2/7 (29%)	3/6 (50%)	3/7 (43%)
NB6	31-05-2020	01-06-2020	1/6 (17%)	4/6 (66%)	4/6 (66%)

<b>NB7</b>	31-05-2020	10-06-2020 – 01-07-2020	8/10 (80%)	NA**	8/10 (80%)
<b>NB8</b>	02-06-2020	03-06-2020	5/10 (50%)	5/9 (56%)	8/10 (80%)
<b>NB9</b>	04-06-2020	07-06-2020	1/7 (14%)	1/7 (14%)	2/7 (29%)
<b>NB10</b>	08-06-2020	11-06-2020	1/8 (13%)	3/8 (38%)	4/8 (50%)
<b>NB11</b>	08-06-2020	11-06-2020	1/3 (33%)	0/2 (0%)	1/3 (33%)
<b>NB12</b>	09-06-2020	11-06-2020	6/9 (66%)	2/8 (25%)	7/9 (78%)
<b>NB13</b>	14-06-2020	11-06-2020 – 18-06-2020	3/3 (33%)	0/2 (0%)	3/3 (33%)
<b>NB14</b>	14-06-2020	14-06-2020	1/3 (100%)	5/6 (83%)	5/6 (83%)
<b>NB15</b>	21-06-2020	10-06-2020 – 30-06-2020	2/2 (100%)	NA**	2/2 (100%)
<b>NB16</b>	21-06-2020	23-06-2020	0/2 (0%)	NA**	0/2 (0%)
<b>Total:</b>			<b>43/88 (49%)</b>	<b>38/75 (51%)</b>	<b>66/97 (68%)</b>

182 \* Serology was done approximately one week before the positive PCR test.

183 \*\* No serology was performed

184

### 185 **Anthropozoonotic transmission of SARS-CoV-2**

186 During the interview on April 28<sup>th</sup>, four out of five employees from NB1 reported that they  
 187 had experienced respiratory symptoms before the outbreak was detected in minks, but none  
 188 of them had been tested for SARS-CoV-2. The first day of symptoms of people working on NB1  
 189 ranged from April 1<sup>st</sup> to May 9<sup>th</sup>. For 16 of the mink, sampled on April 28<sup>th</sup>, and one farm  
 190 employee, sampled on May 4<sup>th</sup>, a WGS was obtained (hCov-  
 191 19/Netherlands/NoordBrabant\_177/2020). The human sequence clusters within the mink  
 192 sequences although it had 7 nucleotides difference with the closest mink sequence (**Figure 1**  
 193 and cluster A in **figures 2 and 3**). On farm NB2, SARS-CoV-2 was diagnosed on April 25<sup>th</sup>.  
 194 Retrospective analysis showed that one employee from NB2 had been hospitalized with SARS-  
 195 CoV-2 on March 31<sup>st</sup>. All samples from the 8 employees taken on April 30<sup>th</sup> were negative by  
 196 RT-PCR but tested positive for SARS-CoV-2 antibodies. The virus sequence obtained from  
 197 animals was distinct from that of farm NB1, indicating a separate introduction (**Figure 2 and**  
 198 **3**, cluster B).

199



217 NB3. Animal and human sequences from farm NB3 were related to those from farm NB1, but  
218 were both part of cluster A.

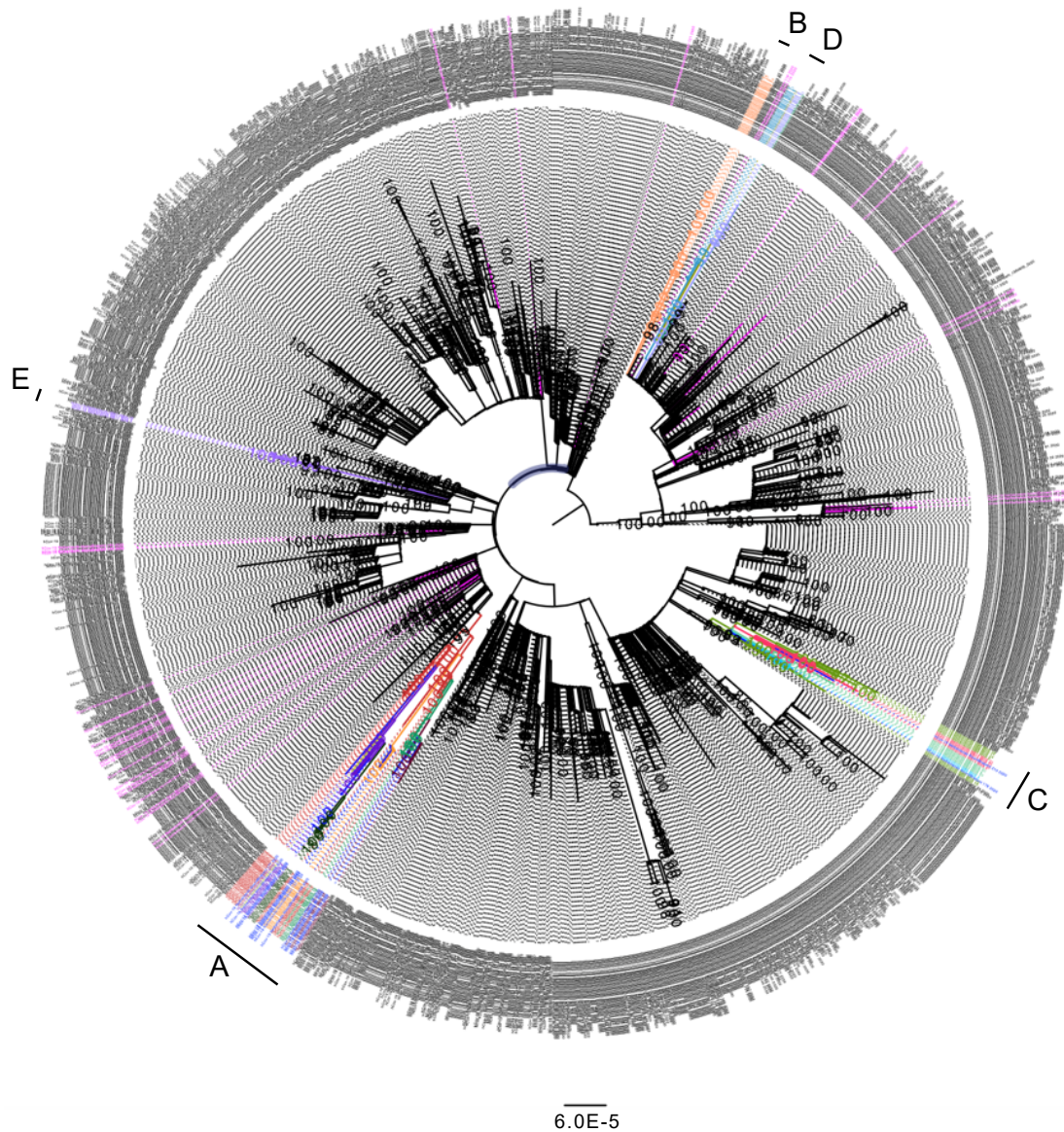
219 Similarly, on mink farm NB7 zoonotic transmission from mink to human most likely  
220 occurred. On this farm, SARS-CoV-2 infection in mink was diagnosed on May 31<sup>st</sup> and  
221 employees initially tested negative for SARS-CoV-2 but started to develop symptoms at a later  
222 stage. Samples were taken between June 10<sup>th</sup> and July 1<sup>st</sup> from 10 employees of which 8 tested  
223 positive for SARS-CoV-2 RNA. From 2 samples WGS could be generated from the employees  
224 which clustered together with the sequences from the animals from this farm.

225

#### 226 **Comparison with national reference database and enhanced regional sampling**

227 The sequences generated from mink farms and from mink farm employees were compared  
228 with the national database consisting of around 1,775 WGS. In addition, to discriminate  
229 between locally acquired infections and mink farm related SARS-CoV-2 infection, and to  
230 determine the potential risk for people living close to mink farms, WGS was also performed  
231 on 34 SARS-CoV-2 positive samples from individuals who live in the same four-digit postal  
232 code area compared to the first four mink farms. These local sequences reflected the general  
233 diversity seen in the Netherlands and were not related to the clusters of mink sequences  
234 found on the mink farms, thereby also giving no indication of spill-over to people living in close  
235 proximity to mink farms (sequences shown in magenta, **Figure 2**). The sequences from the  
236 mink farm investigation were also compared to sequences from Poland (n=65), since many of  
237 the mink farm workers were seasonal migrants from Poland, but these were not related.

238



239

240 **Figure 2: Maximum likelihood analysis of all SARS-CoV-2 Dutch sequences.** The sequences derived  
241 from minks from different farms are indicated with different colors, human sequences related to the  
242 mink farms in blue and samples from similar 4-digit postal code are indicated in magenta. Scale bar  
243 represents units of substitutions per site.

244

#### 245 **Mink farm related sequence clusters**

246 Phylogenetic analysis of the mink SARS-CoV-2 genomes showed that mink sequences of 16  
247 farms grouped into 5 different clusters (**Figure 2 and 3**). Viruses from farms NB1, NB3, NB4,



248 NB8, NB12, NB13 and NB16 belonged to cluster A, sequences from NB2 were a separate  
 249 cluster (B), those from farms NB6, NB7, NB9 and NB14 grouped together in cluster C, NB5,  
 250 NB8, NB10 and NB15 grouped to cluster D, and NB11 had sequences designated as cluster E.  
 251 On farm NB8, SARS-CoV-2 viruses could be found from both cluster A and cluster D. A detailed  
 252 inventory of possible common characteristics, like farm owner, shared personnel, feed  
 253 supplier and veterinary service provider, was made. In some cases, a link was observed with  
 254 the same owners of several farms, for instance for cluster A for NB1 and NB4, and for NB8 and  
 255 NB12. Although NB7, NB11 and NB15 were also linked to the same owner, viruses from these  
 256 farms belonged to cluster C, D and E respectively. No common factor could be identified for  
 257 most farms and clustering could also not be explained by geographic distances as multiple  
 258 clusters were detected in different farms located close to each other (**Table 2 and figure 4**).

259  
 260 **Table 2.** Overview of the clusters detected on the different farms.

Farm:	Date of diagnosis:	Sequence cluster:	Same owner:	Feed supplier:	Vet**:	Number of sequences (human):	Sequence diversity (average):	Mink population size:	Detection***:
NB1	24-04-20	A	NB1, NB4	1	I	17 (1)	0-9 (3.9)	75,711	Notification
NB2	25-04-20	B		1	II	8	0-8 (3.6)	50,473	Notification
NB3	07-05-20	A		2	III	5 (5)	0-2 (0.6)	12,400	Notification
NB4	07-05-20	A	NB1, NB4	1	I	1	NA	67,945	Contact tracing NB1
NB5	31-05-20	D		1	IV	1	NA	38,936	EWS-Ser+PM-1st
NB6	31-05-20	C		3	V	9	0-12 (6.8)	54,515	EWS-Ser+PM-1st
NB7	31-05-20	C	NB7, NB11, NB15	3	II	6 (2)	0-4 (1.4)	79,355	EWS-PM-1st
NB8	02-06-20	A/D	NB8, NB12*	3	V	6 (5)	0-6 (2.6)	39,144	EWS-Ser+PM-1st
NB9	04-06-20	C		2	V	2 (1)	0-3 (1.5)	32,557	EWS-Ser+PM-2nd
NB10	08-06-20	D		3	II	4	0-3 (1.1)	26,824	EWS-Ser+PM-2nd

<b>NB11</b>	08-06-20	E	NB7, NB11, NB15	3	II	4	0-4 (2.2)	38,745	EWS-PM-2nd
<b>NB12</b>	09-06-20	A	NB8, NB12*	3	II	5	0-3 (1.2)	55,352	Notification
<b>NB13</b>	14-06-20	A		3	V	5 (3)	0-5 (3.2)	20,366	EWS-PM-5th
<b>NB14</b>	14-06-20	C		3	II	5 (1)	0-7 (3.7)	28,375	EWS-PM-5th
<b>NB15</b>	21-06-20	D	NB7, NB11, NB15	3	II	5	0-2 (0.6)	35,928	EWS-PM-6th
<b>NB16</b>	21-06-20	A		3	II	5	0-4 (1.6)	66,920	EWS-PM-6th

261 \* There was exchange of personnel in these two locations.

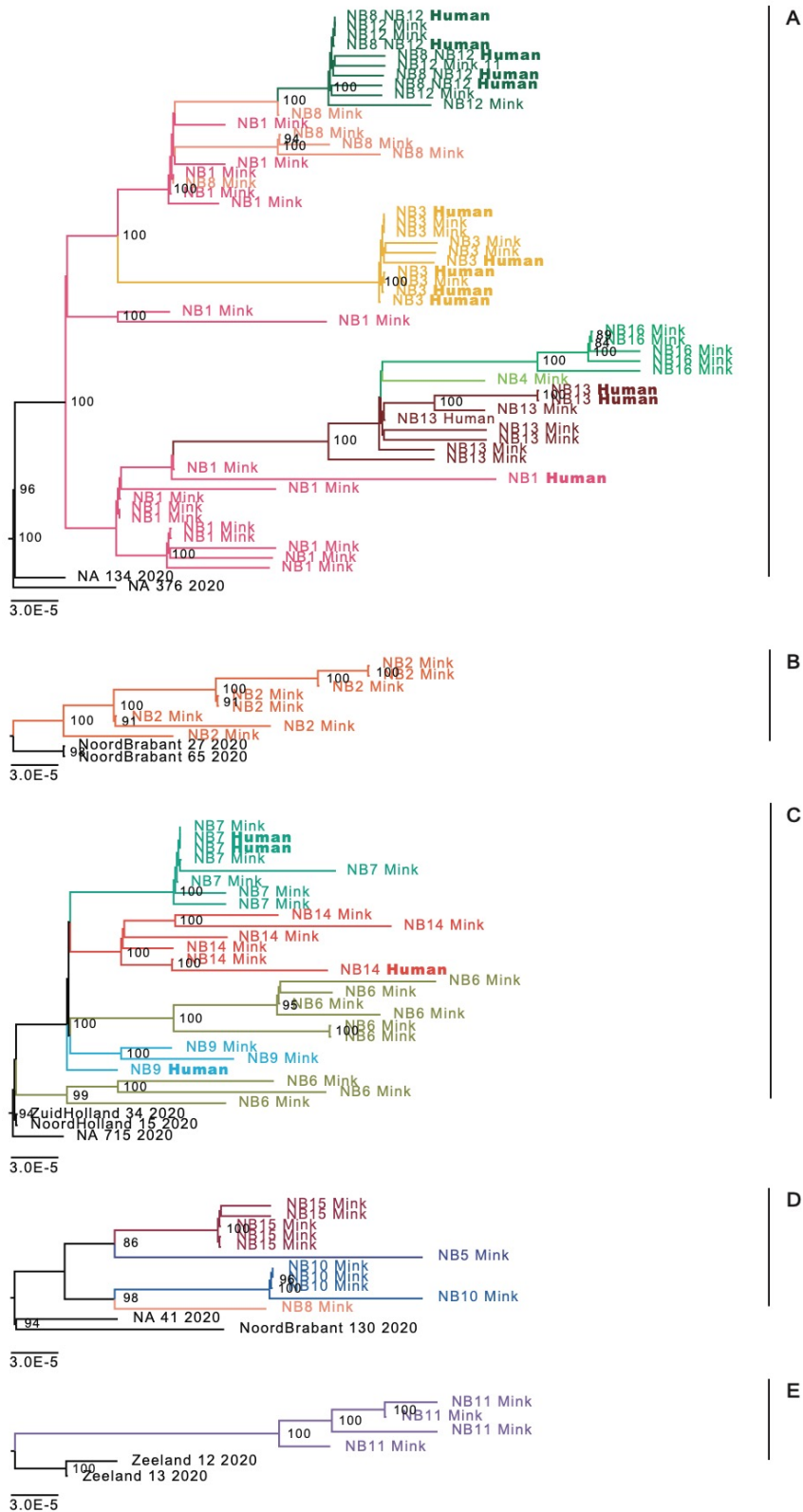
262 \*\* Veterinarian II and V were both from the same veterinary practice.

263 \*\*\* Notification: based on reporting of clinical signs which was obligated from 26 April onwards; EWS-Ser-  
 264 Detection based on a one-off nation-wide compulsory serological screening of all mink farms at the end of  
 265 May/early June by GD Animal Health; EWS-PM-Detection based on the early warning monitoring system  
 266 for which carcasses of animals that died of natural causes were submitted weekly for PCR testing by GD  
 267 Animal Health from the end of May onwards in a weekly cycle (EWS-PM 1<sup>st</sup> to 6<sup>th</sup> post mortem screening).

268

269 In total 18 sequences from mink farm employees or close contacts were generated from seven  
 270 different farms. In most cases, these human sequences were near-identical to the mink  
 271 sequences from the same farm. For NB1 the situation was different and the human sequence  
 272 clusters deeply within the sequences derived from mink (Figure 1), with 7 nucleotides  
 273 difference with the closest related mink sequence. This was also the case on farm NB14, with  
 274 4 nucleotides difference with the closest related mink sequence. Employees sampled at mink  
 275 farm NB8 clustered with animals from NB12 which can be explained by the exchange of  
 276 personnel between these two farms.





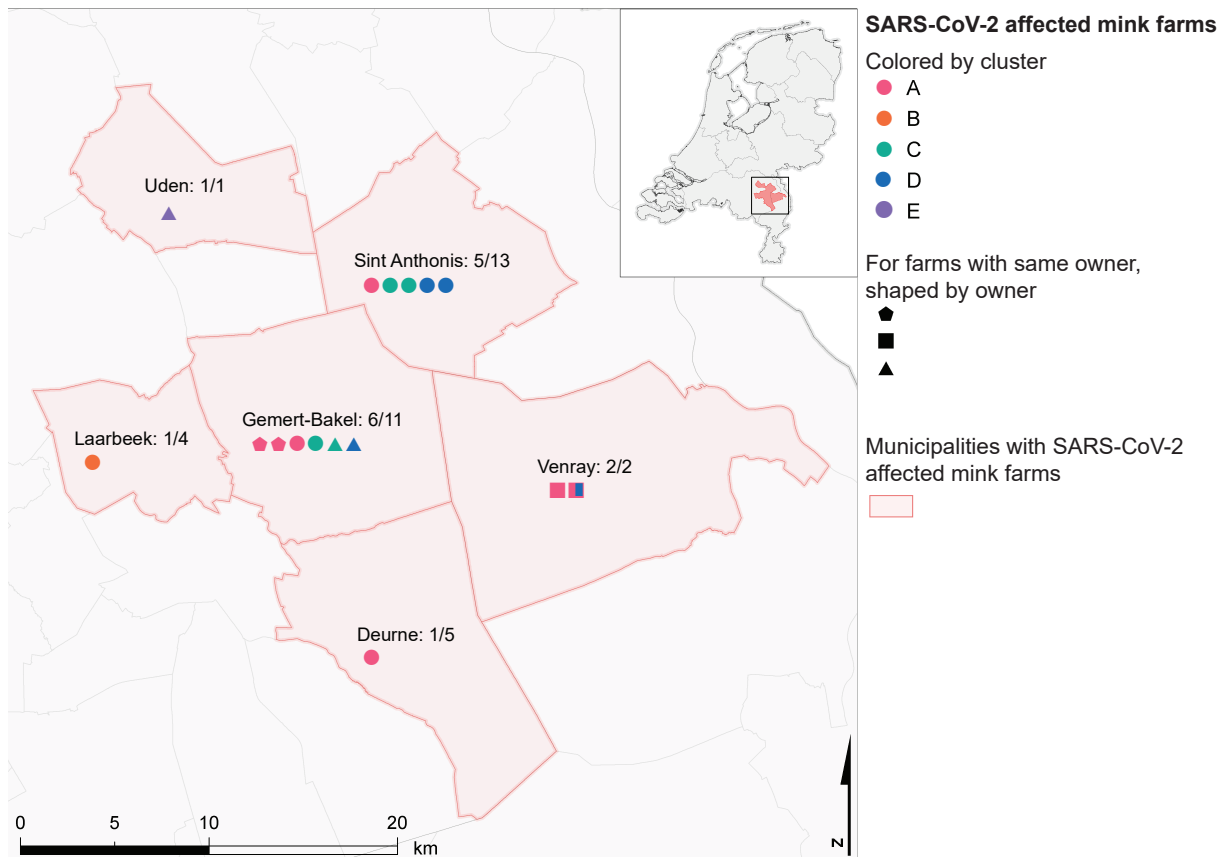
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278

279

280

**Figure 3: Phylogenetic analysis of SARS-CoV-2 strains detected in the 5 mink farm clusters.** The sequences derived from different farms are depicted in different colors. Scale bar represents units of substitutions per site.



281

282 **Figure 4. Geographical overview of SARS-CoV-2 positive mink farms per municipality affected.** The

283 proportion of SARS-CoV-2 positive mink farms over the total number of mink farms (CBS, 2019) is

284 indicated. Symbols for positive farms are colored by cluster and shapes indicate farms with a same

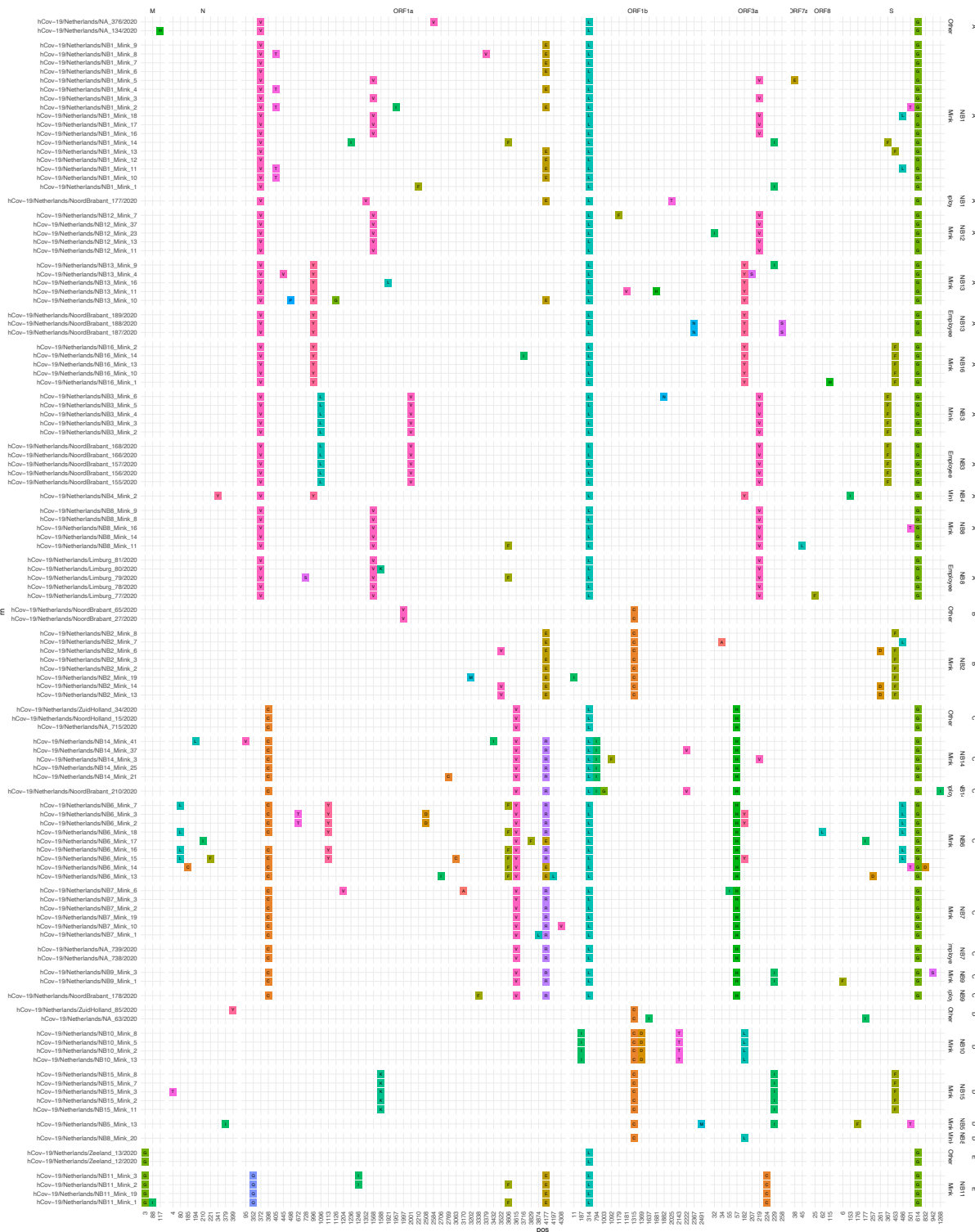
285 owner.

286

287 **Within farm diversity**

288 SARS-CoV-2 was detected on mink farm NB1-NB4 after reports of respiratory symptoms and  
289 increased mortality in mink. The sequences from farm NB1 had between 0 and 9 nucleotides  
290 differences (average 3.9 nucleotides) and from NB2 between 0 and 8 nucleotides differences  
291 (average of 3.6), which is much more than what has been observed in outbreaks in human  
292 settings. The sequences from NB3 had 0 to 2 nucleotides difference suggesting that the virus  
293 was recently introduced, in line with the observed disease in humans, which occurred in the  
294 weeks post diagnosis of the infection in mink. After the initial detection of SARS-CoV-2 on  
295 mink farms, farms were screened weekly. The first, second, fifth and sixth weekly screening  
296 yielded new positives. The sequences of mink at NB6 had between 0 and 12 nucleotides  
297 differences, whereas diversity was lower for the subsequent farm sequences (Table 2).

298         Several non-synonymous mutations were identified among the mink sequences  
299 compared to the Wuhan reference sequence NC\_045512.2. However, no particular amino  
300 acid substitutions were found in all mink samples (**Figure 5**). Of note, three of the clusters had  
301 the position 614G variant (clusters A, C and E), and 2 had the original variant. There were no  
302 obvious differences in the presentation of disease in animals or humans between clusters  
303 based on the data available at this stage, but further data collection and analysis, also for cases  
304 after NB16, are ongoing to investigate this further. The observed mutations can also be found  
305 in the general population and the same mutations also were found in human cases which were  
306 related to the mink farms.



307  
 308 **Figure 5. Overview of the specific amino acid mutations found in mink farms.** Above the x-axis the  
 309 open reading frames (ORF) are indicated and on the x-axis the amino acid position within each  
 310 ORF is indicated. On the y-axis the sequence names are indicated and on the right side of the

311 graph the cluster numbers and specific farm identifiers and the type of host are used to group  
312 the samples.

313

## 314 **Discussion**

315 Here we show ongoing SARS-CoV-2 transmission in mink farms and spill-over events to  
316 humans. To the best of our knowledge, these are the first animal to human SARS-CoV-2  
317 transmission events documented. More research in minks and other mustelid species, to  
318 demonstrate if these species can be a true reservoir of SARS-CoV-2 although from our  
319 observations we consider this likely. After the detection of SARS-CoV-2 on mink farms, 68% of  
320 the tested farm workers and/or relatives or contacts were shown to be infected with SARS-  
321 CoV-2, indicating that contact with SARS-CoV-2 infected mink is a risk factor for contracting  
322 COVID-19.

323         A high diversity in the sequences from some mink farms was observed which most  
324 likely can be explained by many generations of infected animals before an increase in  
325 mortality was observed. The current estimates are that the substitution rate of SARS-CoV-2 is  
326 around  $1.16 \times 10^{-3}$  substitutions/site/year (53), which corresponds to around one mutation  
327 per two weeks. This could mean that the virus was already circulating in mink farms for some  
328 time. However, there was also a relatively high sequence diversity observed in farms which  
329 still tested negative one week prior, hinting towards a faster evolution of the virus in the mink  
330 population. This can indicate that the virus might replicate more efficiently in mink or might  
331 have acquired mutations which makes the virus more virulent. However, no specific mutations  
332 were found in all mink samples, making increased virulence less likely. In addition, mink farms  
333 have large populations of animals which could lead to very efficient virus transmission.  
334 Generation intervals for SARS-CoV-2 in humans have been estimated to be around 4-5

335 days(54), but with high dose exposure in a high-density farm could potentially be shorter.  
336 Recently, a specific mutation in the spike protein (D614G) was shown to result in an increased  
337 virulence *in vitro* (55), while it was not associated an increased growth rate for cluster nor an  
338 increased mortality (56). This mutation was present in farm clusters A, C and E, but no obvious  
339 differences in clinical presentation, disease severity, or rate of transmission to humans was  
340 observed.

341         While we found sequences matching with the animal sequences on several farms, not  
342 all of these can be considered direct zoonotic transmissions. For instance, the two employees  
343 from mink farm NB3 were most likely infected while working at the mink farm given the  
344 specific clustering in the phylogenetic tree and the timing of infection. Subsequent human  
345 infections may have originated from additional zoonotic infections, or from human to human  
346 transmission within their household. Further proof that animals were the most likely source  
347 of infection was provided by the clear phylogenetic separation between farm related human  
348 cases and animal cases, from sequences from cases within the same 4-digit postal code area.  
349 Spill-back into the community living in the same 4-digit postal code area was not observed  
350 using sequence data, but cannot be entirely ruled out as the testing strategy during April and  
351 May was focusing on health care workers, persons with more severe symptoms, and persons  
352 at risk for complications, rather than monitoring community transmission and milder cases.

353         While the number of SARS-CoV-2 infected individuals was decreasing in the  
354 Netherlands in May and June, an increase in detection of SARS-CoV-2 in mink farms was  
355 observed. Based on WGS these sequences are part of multiple individual transmission chains  
356 linked to the mink farms and are not a reflection of the situation in the human population  
357 during this time. In some cases, the farms had the same owner but in other cases no  
358 epidemiological link could be established. People coming to the different farms might be a

359 source but also semi-wild cats roaming around the farms or wildlife might play a role (27). So  
360 far, the investigation failed to identify common factors that might explain farm to farm spread.  
361 During interviews, it became clear that farms had occasionally hired temporary workers that  
362 had not been included in the testing and were lost to follow-up, stressing the need for vigorous  
363 biosecurity and occupational health guidance. Since our observation, SARS-CoV-2 infections  
364 have also been described in mink farms in Denmark, Spain and the USA (57–59), and mink  
365 farming is common in other regions of the world as well, also in China where around 26 million  
366 mink pelts are produced on a yearly basis (60). The population size and the structure of mink  
367 farms is such that it is conceivable that SARS-CoV-2 – once introduced – could continue to  
368 circulate. Therefore, continued monitoring and cooperation between human and animal  
369 health services is crucial to prevent the animals serving as a reservoir for continued infection  
370 in humans.

371

## 372 **References**

- 373 1. N. Zhu, D. Zhang, W. Wang, X. Li, B. Yang, J. Song, X. Zhao, B. Huang, W. Shi, R. Lu, P.  
374 Niu, F. Zhan, X. Ma, D. Wang, W. Xu, G. Wu, G. F. Gao, W. Tan, A novel coronavirus from  
375 patients with pneumonia in China, 2019. *N. Engl. J. Med.* **382**, 727–733 (2020).
- 376 2. E. Dong, H. Du, L. Gardner, An interactive web-based dashboard to track COVID-19 in  
377 real time. *Lancet Infect. Dis.* **0** (2020), , doi:10.1016/S1473-3099(20)30120-1.
- 378 3. B. B. Oude Munnink, D. F. Nieuwenhuijse, M. Stein, Á. O’Toole, M. Haverkate, M.  
379 Mollers, S. K. Kamga, C. Schapendonk, M. Pronk, P. Lexmond, A. van der Linden, T.  
380 Bestebroer, I. Chestakova, R. J. Overmars, S. van Nieuwkoop, R. Molenkamp, A. A. van  
381 der Eijk, C. GeurtsvanKessel, H. Vennema, A. Meijer, A. Rambaut, J. van Dissel, R. S.  
382 Sikkema, A. Timen, M. Koopmans, G. J. A. P. M. Oudehuis, J. Schinkel, J. Kluytmans, M.

- 383 Kluytmans-van den Bergh, W. van den Bijllaardt, R. G. Berntvelsen, M. M. L. van Rijen,  
384 P. Schneeberger, S. Pas, B. M. Diederer, A. M. C. Bergmans, P. A. V. van der Eijk, J.  
385 Verweij, A. G. N. Buiting, R. Streefkerk, A. P. Aldenkamp, P. de Man, J. G. M. Koelemal,  
386 D. Ong, S. Paltansing, N. Veassen, J. Slevén, L. Bakker, H. Brockhoff, A. Rietveld, F.  
387 Slijkerman Megelink, J. Cohen Stuart, A. de Vries, W. van der Reijden, A. Ros, E. Lodder,  
388 E. Verspui-van der Eijk, I. Huijskens, E. M. Kraan, M. P. M. van der Linden, S. B. Debast,  
389 N. Al Naiemi, A. C. M. Kroes, M. Damen, S. Dinant, S. Lekkerkerk, O. Pontesilli, P. Smit,  
390 C. van Tienen, P. C. R. Godschalk, J. van Pelt, A. Ott, C. van der Weijden, H. Wertheim,  
391 J. Rahamat-Langendoen, J. Reimerink, R. Bodewes, E. Duizer, B. van der Veer, C.  
392 Reusken, S. Lutgens, P. Schneeberger, M. Hermans, P. Wever, A. Leenders, H. ter  
393 Waarbeek, C. Hoebe, Rapid SARS-CoV-2 whole-genome sequencing and analysis for  
394 informed public health decision-making in the Netherlands. *Nat. Med.*, 1–6 (2020).
- 395 4. F. Wu, S. Zhao, B. Yu, Y. M. Chen, W. Wang, Z. G. Song, Y. Hu, Z. W. Tao, J. H. Tian, Y. Y.  
396 Pei, M. L. Yuan, Y. L. Zhang, F. H. Dai, Y. Liu, Q. M. Wang, J. J. Zheng, L. Xu, E. C. Holmes,  
397 Y. Z. Zhang, A new coronavirus associated with human respiratory disease in China.  
398 *Nature*. **579**, 265–269 (2020).
- 399 5. H. Zhou, X. Chen, T. Hu, J. Li, H. Song, Y. Liu, P. Wang, D. Liu, J. Yang, E. C. Holmes, A. C.  
400 Hughes, Y. Bi, W. Shi, A Novel Bat Coronavirus Closely Related to SARS-CoV-2 Contains  
401 Natural Insertions at the S1/S2 Cleavage Site of the Spike Protein. *Curr. Biol.* (2020),  
402 doi:10.1016/j.cub.2020.05.023.
- 403 6. P. Zhou, X. Lou Yang, X. G. Wang, B. Hu, L. Zhang, W. Zhang, H. R. Si, Y. Zhu, B. Li, C. L.  
404 Huang, H. D. Chen, J. Chen, Y. Luo, H. Guo, R. Di Jiang, M. Q. Liu, Y. Chen, X. R. Shen, X.  
405 Wang, X. S. Zheng, K. Zhao, Q. J. Chen, F. Deng, L. L. Liu, B. Yan, F. X. Zhan, Y. Y. Wang,  
406 G. F. Xiao, Z. L. Shi, A pneumonia outbreak associated with a new coronavirus of



- 407 probable bat origin. *Nature*. **579**, 270–273 (2020).
- 408 7. T. T. Y. Lam, M. H. H. Shum, H. C. Zhu, Y. G. Tong, X. B. Ni, Y. S. Liao, W. Wei, W. Y. M.  
409 Cheung, W. J. Li, L. F. Li, G. M. Leung, E. C. Holmes, Y. L. Hu, Y. Guan, Identifying SARS-  
410 CoV-2 related coronaviruses in Malayan pangolins. *Nature*, 1–6 (2020).
- 411 8. G. Z. Han, Pangolins Harbor SARS-CoV-2-Related Coronaviruses. *Trends Microbiol.*  
412 (2020), , doi:10.1016/j.tim.2020.04.001.
- 413 9. M. F. Boni, P. Lemey, X. Jiang, T. T.-Y. Lam, B. Perry, T. Castoe, A. Rambaut, D. L.  
414 Robertson, *bioRxiv*, in press, doi:10.1101/2020.03.30.015008.
- 415 10. J. Shi, Z. Wen, G. Zhong, H. Yang, C. Wang, B. Huang, R. Liu, X. He, L. Shuai, Z. Sun, Y.  
416 Zhao, P. Liu, L. Liang, P. Cui, J. Wang, X. Zhang, Y. Guan, W. Tan, G. Wu, H. Chen, Z. Bu,  
417 Susceptibility of ferrets, cats, dogs, and other domesticated animals to SARS–  
418 coronavirus 2. *Science* (80-. ), eabb7015 (2020).
- 419 11. P. J. Halfmann, M. Hatta, S. Chiba, T. Maemura, S. Fan, M. Takeda, N. Kinoshita, S.-I.  
420 Hattori, Y. Sakai-Tagawa, K. Iwatsuki-Horimoto, M. Imai, Y. Kawaoka, Transmission of  
421 SARS-CoV-2 in Domestic Cats. *N. Engl. J. Med.*, NEJMc2013400 (2020).
- 422 12. M. Richard, A. Kok, D. de Meulder, T. M. Bestebroer, M. M. Lamers, N. M. A. Okba, M.  
423 F. van Vliissingen, B. Rockx, B. L. Haagmans, M. P. G. Koopmans, R. A. M. Fouchier, S.  
424 Herfst, *bioRxiv*, in press, doi:10.1101/2020.04.16.044503.
- 425 13. S. F. Sia, L.-M. Yan, K. Fung, J. M. Nicholls, M. Peiris, H.-L. Yen, Pathogenesis and  
426 transmission of SARS-CoV-2 virus in golden Syrian hamsters SUBJECT AREAS Infectious  
427 Diseases Small Animal Medicine (2020), doi:10.21203/rs.3.rs-20774/v1.
- 428 14. J. F. W. Chan, A. J. Zhang, S. Yuan, V. K. M. Poon, C. C. S. Chan, A. C. Y. Lee, W. M. Chan,  
429 Z. Fan, H. W. Tsoi, L. Wen, R. Liang, J. Cao, Y. Chen, K. Tang, C. Luo, J. P. Cai, K. H. Kok,  
430 H. Chu, K. H. Chan, S. Sridhar, Z. Chen, H. Chen, K. K. W. To, K. Y. Yuen, Simulation of the

- 431 clinical and pathological manifestations of Coronavirus Disease 2019 (COVID-19) in  
432 golden Syrian hamster model: implications for disease pathogenesis and  
433 transmissibility. *Clin. Infect. Dis.* (2020), doi:10.1093/cid/ciaa325.
- 434 15. V. J. Munster, F. Feldmann, B. N. Williamson, N. van Doremalen, L. Pérez-Pérez, J.  
435 Schulz, K. Meade-White, A. Okumura, J. Callison, B. Brumbaugh, V. A. Avanzato, R.  
436 Rosenke, P. W. Hanley, G. Saturday, D. Scott, E. R. Fischer, E. de Wit, Respiratory disease  
437 in rhesus macaques inoculated with SARS-CoV-2. *Nature*, 1–7 (2020).
- 438 16. Y. Zhao, J. Wang, D. Kuang, J. Xu, M. Yang, C. Ma, S. Zhao, J. Li, H. Long, K. Ding, J. Gao,  
439 J. Liu, H. Wang, H. Li, Y. Yang, W. Yu, J. Yang, Y. Zheng, D. Wu, S. Lu, H. Liu, X. Peng,  
440 *bioRxiv*, in press, doi:10.1101/2020.04.30.029736.
- 441 17. B. Rockx, T. Kuiken, S. Herfst, T. Bestebroer, M. M. Lamers, B. B. Oude Munnink, D. de  
442 Meulder, G. van Amerongen, J. van den Brand, N. M. A. Okba, D. Schipper, P. van Run,  
443 L. Leijten, R. Sikkema, E. Verschoor, B. Verstrepen, W. Bogers, J. Langermans, C.  
444 Drosten, M. Fentener van Vlissingen, R. Fouchier, R. de Swart, M. Koopmans, B. L.  
445 Haagmans, Comparative pathogenesis of COVID-19, MERS, and SARS in a nonhuman  
446 primate model. *Science (80-. )*, eabb7314 (2020).
- 447 18. C. Woolsey, V. Borisevich, A. N. Prasad, K. N. Agans, D. J. Deer, N. S. Dobias, J. C.  
448 Heymann, S. L. Foster, C. B. Levine, L. Medina, K. Melody, J. B. Geisbert, K. A. Fenton, T.  
449 W. Geisbert, R. W. Cross, *bioRxiv Prepr. Serv. Biol.*, in press,  
450 doi:10.1101/2020.05.17.100289.
- 451 19. S. Lu, Y. Zhao, W. Yu, Y. Yang, J. Gao, J. Wang, D. Kuang, M. Yang, J. Yang, C. Ma, J. Xu,  
452 H. Li, S. Zhao, J. Li, H. Wang, H. Long, J. Zhou, F. Luo, K. Ding, D. Wu, Y. Zhang, Y. Dong,  
453 Y. Liu, Y. Zheng, X. Lin, L. Jiao, H. Zheng, Q. Dai, Q. Sun, Y. Hu, C. Ke, H. Liu, X. Peng,  
454 *bioRxiv*, in press, doi:10.1101/2020.04.08.031807.

- 455 20. B. L. Haagmans, D. Noack, N. M. Okba, W. Li, C. Wang, R. de Vries, S. Herfst, D. de  
456 Meulder, P. van Run, B. Rijnders, C. Rokx, F. van Kuppeveld, F. Grosveld, C.  
457 GeurtsvanKessel, M. Koopmans, B. Jan Bosch, T. Kuiken, B. Rockx, *bioRxiv*, in press,  
458 doi:10.1101/2020.08.24.264630.
- 459 21. K. Schlottau, M. Rissmann, A. Graaf, J. Schön, J. Sehl, C. Wylezich, D. Höper, T. C.  
460 Mettenleiter, A. Balkema-Buschmann, T. Harder, C. Grund, D. Hoffmann, A. Breithaupt,  
461 M. Beer, Experimental Transmission Studies of SARS-CoV-2 in Fruit Bats, Ferrets, Pigs  
462 and Chickens. *SSRN Electron. J.* (2020), doi:10.2139/ssrn.3578792.
- 463 22. D. L. Suarez, M. J. Pantin-Jackwood, D. E. Swayne, S. A. Lee, S. M. Deblois, E. Spackman,  
464 *bioRxiv*, in press, doi:10.1101/2020.06.16.154658.
- 465 23. T. H. C. Sit, C. J. Brackman, S. M. Ip, K. W. S. Tam, P. Y. T. Law, E. M. W. To, V. Y. T. Yu, L.  
466 D. Sims, D. N. C. Tsang, D. K. W. Chu, R. A. P. M. Perera, L. L. M. Poon, M. Peiris, Infection  
467 of dogs with SARS-CoV-2. *Nature*, 1–6 (2020).
- 468 24. C. Sailleau, M. Dumarest, J. Vanhomwegen, M. Delaplace, V. Caro, A. Kwasiborski, V.  
469 Hourdel, P. Chevaillier, A. Barbarino, L. Comtet, P. Pourquier, B. Klonjkowski, J. C.  
470 Manuguerra, S. Zientara, S. Le Poder, First detection and genome sequencing of SARS-  
471 CoV-2 in an infected cat in France. *Transbound. Emerg. Dis.* (2020),  
472 doi:10.1111/tbed.13659.
- 473 25. A. Newman, D. Smith, R. R. Ghai, R. M. Wallace, M. K. Torchetti, C. Loiacono, L. S.  
474 Murrell, A. Carpenter, S. Moroff, J. A. Rooney, C. Barton Behravesh, First Reported  
475 Cases of SARS-CoV-2 Infection in Companion Animals - New York, March-April 2020.  
476 *MMWR. Morb. Mortal. Wkly. Rep.* **69**, 710–713 (2020).
- 477 26. Promed Post – ProMED-mail, (available at [https://promedmail.org/promed-](https://promedmail.org/promed-post/?id=7314521)  
478 [post/?id=7314521](https://promedmail.org/promed-post/?id=7314521)).

- 479 27. N. Oreshkova, R.-J. Molenaar, S. Vreman, F. Harders, B. B. O. Munnink, R. Hakze, N.  
480 Gerhards, P. Tolsma, R. Bouwstra, R. Sikkema, M. Tacken, M. M. T. de Rooij, E.  
481 Weesendorp, M. Engelsma, C. Brusckke, L. A. M. Smit, M. Koopmans, W. H. M. van der  
482 Poel, A. Stegeman, *bioRxiv*, in press, doi:10.1101/2020.05.18.101493.
- 483 28. R. Gollakner, I. Capua, Is COVID-19 the first pandemic that evolves into a panzootic?  
484 *Vet. Ital.* **56** (2020), doi:10.12834/VetIt.2246.12523.1.
- 485 29. N. Oreshkova, R. J. Molenaar, S. Vreman, F. Harders, B. B. Oude Munnink, R. W. Hakze-  
486 van der Honing, N. Gerhards, P. Tolsma, R. Bouwstra, R. S. Sikkema, M. G. Tacken, M.  
487 M. de Rooij, E. Weesendorp, M. Y. Engelsma, C. J. Brusckke, L. A. Smit, M. Koopmans,  
488 W. H. van der Poel, A. Stegeman, SARS-CoV-2 infection in farmed minks, the  
489 Netherlands, April and May 2020. *Eurosurveillance.* **25**, 2001005 (2020).
- 490 30. Q. Zhang, H. Zhang, K. Huang, Y. Yang, X. Hui, J. Gao, X. He, C. Li, W. Gong, Y. Zhang, C.  
491 Peng, X. Gao, H. Chen, Z. Zou, Z. Shi, M. Jin, *bioRxiv*, in press,  
492 doi:10.1101/2020.04.01.021196.
- 493 31. E. I. Patterson, G. Elia, A. Grassi, A. Giordano, C. Desario, M. Medardo, S. L. Smith, E. R.  
494 Anderson, E. Lorusso, M. S. Lucente, G. Lanave, S. Lauzi, U. Bonfanti, A. Stranieri, V.  
495 Martella, Fabrizio Solari Basano, V. R. Barrs, A. D. Radford, G. L. Hughes, S. Paltrinieri,  
496 N. Decaro, *bioRxiv*, in press, doi:10.1101/2020.07.21.214346.
- 497 32. R. J. Molenaar, S. Vreman, R. W. Hakze-van der Honing, R. Zwart, J. de Rond, E.  
498 Weesendorp, L. A. M. Smit, M. Koopmans, R. Bouwstra, A. Stegeman, W. H. M. van der  
499 Poel, Clinical and Pathological Findings in SARS-CoV-2 Disease Outbreaks in Farmed  
500 Mink (*Neovison vison*). *Vet. Pathol.* (2020), doi:10.1177/0300985820943535.
- 501 33. Bedrijfsmatig gehouden dieren en SARS-CoV-2 | Nieuws en media | NVWA, (available  
502 at <https://www.nvwa.nl/nieuws-en-media/actuele->

- 503 onderwerpen/corona/g/bedrijfsmatig-gehouden-dieren-en-corona).
- 504 34. B. B. Oude Munnink, E. Münger, D. F. Nieuwenhuijse, R. Kohl, A. van der Linden, C. M.  
505 E. Schapendonk, H. van der Jeugd, M. Kik, J. M. Rijks, C. B. E. M. Reusken, M. Koopmans,  
506 Genomic monitoring to understand the emergence and spread of Usutu virus in the  
507 Netherlands, 2016-2018. *Sci. Rep.* **10**, 2798 (2020).
- 508 35. A. Arias, S. J. Watson, D. Asogun, E. A. Tobin, J. Lu, M. V. T. Phan, U. Jah, R. E. G.  
509 Wadoun, L. Meredith, L. Thorne, S. Caddy, A. Tarawalie, P. Langat, G. Dudas, N. R. Faria,  
510 S. Dellicour, A. Kamara, B. Kargbo, B. O. Kamara, S. Gevao, D. Cooper, M. Newport, P.  
511 Horby, J. Dunning, F. Sahr, T. Brooks, A. J. H. Simpson, E. GropPELLI, G. Liu, N. Mulakken,  
512 K. Rhodes, J. Akpablie, Z. Yoti, M. Lamunu, E. Vitto, P. Otim, C. Owilli, I. Boateng, L.  
513 Okoror, E. Omomoh, J. Oyakhilome, R. Omiunu, I. Yemisis, D. Adomeh, S.  
514 Ehikhiametalor, P. Akhilomen, C. Aire, A. Kurth, N. Cook, J. Baumann, M. Gabriel, R.  
515 Wölfel, A. Di Caro, M. W. Carroll, S. Günther, J. Redd, D. Naidoo, O. G. Pybus, A.  
516 Rambaut, P. Kellam, I. Goodfellow, M. Cotten, Rapid outbreak sequencing of Ebola virus  
517 in Sierra Leone identifies transmission chains linked to sporadic cases. *Virus Evol.* **2**,  
518 vew016 (2016).
- 519 36. N. R. Faria, M. U. G. Kraemer, S. C. Hill, J. G. de Jesus, R. S. Aguiar, F. C. M. Iani, J. Xavier,  
520 J. Quick, L. du Plessis, S. Dellicour, J. Théz , R. D. O. Carvalho, G. Baele, C.-H. Wu, P. P.  
521 Silveira, M. B. Arruda, M. A. Pereira, G. C. Pereira, J. Lourenço, U. Obolski, L. Abade, T.  
522 I. Vasylyeva, M. Giovanetti, D. Yi, D. J. Weiss, G. R. W. Wint, F. M. Shearer, S. Funk, B.  
523 Nikolay, V. Fonseca, T. E. R. Adelino, M. A. A. Oliveira, M. V. F. Silva, L. Sacchetto, P. O.  
524 Figueiredo, I. M. Rezende, E. M. Mello, R. F. C. Said, D. A. Santos, M. L. Ferraz, M. G.  
525 Brito, L. F. Santana, M. T. Menezes, R. M. Brindeiro, A. Tanuri, F. C. P. dos Santos, M. S.  
526 Cunha, J. S. Nogueira, I. M. Rocco, A. C. da Costa, S. C. V. Komninakis, V. Azevedo, A. O.

- 527 Chieppe, E. S. M. Araujo, M. C. L. Mendonça, C. C. dos Santos, C. D. dos Santos, A. M.  
528 Mares-Guia, R. M. R. Nogueira, P. C. Sequeira, R. G. Abreu, M. H. O. Garcia, A. L. Abreu,  
529 O. Okumoto, E. G. Kroon, C. F. C. de Albuquerque, K. Lewandowski, S. T. Pullan, M.  
530 Carroll, T. de Oliveira, E. C. Sabino, R. P. Souza, M. A. Suchard, P. Lemey, G. S. Trindade,  
531 B. P. Drumond, A. M. B. Filippis, N. J. Loman, S. Cauchemez, L. C. J. Alcantara, O. G.  
532 Pybus, Genomic and epidemiological monitoring of yellow fever virus transmission  
533 potential. *Science (80-. )*. **361**, 894–899 (2018).
- 534 37. J. Quick, N. J. Loman, S. Duraffour, J. T. Simpson, E. Severi, L. Cowley, J. A. Bore, R.  
535 Koundouno, G. Dudas, A. Mikhail, N. Ouédraogo, B. Afrough, A. Bah, J. H. J. Baum, B.  
536 Becker-Ziaja, J. P. Boettcher, M. Cabeza-Cabrerizo, Á. Camino-Sánchez, L. L. Carter, J.  
537 Doerrbecker, T. Enkirch, I. G.- Dorival, N. Hetzelt, J. Hinzmann, T. Holm, L. E.  
538 Kafetzopoulou, M. Koropogui, A. Kosgey, E. Kuisma, C. H. Logue, A. Mazzarelli, S. Meisel,  
539 M. Mertens, J. Michel, D. Ngabo, K. Nitzsche, E. Pallasch, L. V. Patrono, J. Portmann, J.  
540 G. Repits, N. Y. Rickett, A. Sachse, K. Singethan, I. Vitoriano, R. L. Yemanaberhan, E. G.  
541 Zekeng, T. Racine, A. Bello, A. A. Sall, O. Faye, O. Faye, N. Magassouba, C. V. Williams,  
542 V. Amburgey, L. Winona, E. Davis, J. Gerlach, F. Washington, V. Monteil, M. Jourdain,  
543 M. Bererd, A. Camara, H. Somlare, A. Camara, M. Gerard, G. Bado, B. Baillet, D. Delaune,  
544 K. Y. Nebie, A. Diarra, Y. Savane, R. B. Pallawo, G. J. Gutierrez, N. Milhano, I. Roger, C. J.  
545 Williams, F. Yattara, K. Lewandowski, J. Taylor, P. Rachwal, D. J. Turner, G. Pollakis, J. A.  
546 Hiscox, D. A. Matthews, M. K. O. Shea, A. M. Johnston, D. Wilson, E. Hutley, E. Smit, A.  
547 Di Caro, R. Wölfel, K. Stoecker, E. Fleischmann, M. Gabriel, S. A. Weller, L. Koivogui, B.  
548 Diallo, S. Keïta, A. Rambaut, P. Formenty, S. Günther, M. W. Carroll, Real-time, portable  
549 genome sequencing for Ebola surveillance. *Nature*. **530**, 228–232 (2016).
- 550 38. R. S. Sikkema, S. D. Pas, D. F. Nieuwenhuijse, Á. O’Toole, J. Verweij, A. van der Linden,

- 551 I. Chestakova, C. Schapendonk, M. Pronk, P. Lexmond, T. Bestebroer, R. J. Overmars, S.  
552 van Nieuwkoop, W. van den Bijllaardt, R. G. Bentvelsen, M. M. L. van Rijen, A. G. M.  
553 Buiting, A. J. G. van Oudheusden, B. M. Diederer, A. M. C. Bergmans, A. van der Eijk, R.  
554 Molenkamp, A. Rambaut, A. Timen, J. A. J. W. Kluytmans, B. B. Oude Munnink, M. F. Q.  
555 Kluytmans van den Bergh, M. P. G. Koopmans, COVID-19 in health-care workers in three  
556 hospitals in the south of the Netherlands: a cross-sectional study. *Lancet Infect. Dis.* **0**  
557 (2020), doi:10.1016/S1473-3099(20)30527-2.
- 558 39. RIVM, Signaleringsoverleg zoönosen | RIVM, (available at  
559 [https://www.rivm.nl/surveillance-van-infectieziekten/signalering-](https://www.rivm.nl/surveillance-van-infectieziekten/signalering-infectieziekten/signaleringsoverleg-zoonosen)  
560 [infectieziekten/signaleringsoverleg-zoonosen](https://www.rivm.nl/surveillance-van-infectieziekten/signaleringsoverleg-zoonosen)).
- 561 40. RIVM, Outbreak Management Team (OMT) | RIVM, (available at  
562 <https://www.rivm.nl/coronavirus-covid-19/omt>).
- 563 41. A. Kroneman, H. Vennema, K. Deforche, H. v d Avoort, S. Penaranda, M. S. Oberste, J.  
564 Vinje, M. Koopmans, An automated genotyping tool for enteroviruses and noroviruses.  
565 *J Clin Virol.* **51**, 121–125 (2011).
- 566 42. V. M. Corman, O. Landt, M. Kaiser, R. Molenkamp, A. Meijer, D. K. Chu, T. Bleicker, S.  
567 Brünink, J. Schneider, M. L. Schmidt, D. G. Mulders, B. L. Haagmans, B. van der Veer, S.  
568 van den Brink, L. Wijsman, G. Goderski, J.-L. Romette, J. Ellis, M. Zambon, M. Peiris, H.  
569 Goossens, C. Reusken, M. P. Koopmans, C. Drosten, Detection of 2019 novel  
570 coronavirus (2019-nCoV) by real-time RT-PCR. *Eurosurveillance.* **25**, 2000045 (2020).
- 571 43. C. H. GeurtsvanKessel, N. M. A. Okba, Z. Igloi, S. Bogers, C. W. E. Embregts, B. M.  
572 Laksono, L. Leijten, C. Rokx, B. Rijnders, J. Rahamat-Langendoen, J. P. C. van den Akker,  
573 J. J. A. van Kampen, A. A. van der Eijk, R. S. van Binnendijk, B. Haagmans, M. Koopmans,  
574 An evaluation of COVID-19 serological assays informs future diagnostics and exposure

- 575 assessment. *Nat. Commun.* **11**, 1–5 (2020).
- 576 44. B. B. Oude Munnink, D. F. Nieuwenhuijse, R. S. Sikkema, M. Koopmans, Validating  
577 Whole Genome Nanopore Sequencing, using Usutu Virus as an Example. *J. Vis. Exp.*,  
578 e60906 (2020).
- 579 45. R. C. Edgar, MUSCLE: multiple sequence alignment with high accuracy and high  
580 throughput. *Nucleic Acids Res.* **32**, 1792–1797 (2004).
- 581 46. L. T. Nguyen, H. A. Schmidt, A. von Haeseler, B. Q. Minh, IQ-TREE: a fast and effective  
582 stochastic algorithm for estimating maximum-likelihood phylogenies. *Mol Biol Evol.* **32**,  
583 268–274 (2015).
- 584 47. Y. Shu, J. McCauley, GISAID: Global initiative on sharing all influenza data – from vision  
585 to reality. *Eurosurveillance.* **22** (2017), , doi:10.2807/1560-7917.ES.2017.22.13.30494.
- 586 48. H. Wickham, *ggplot2: Elegant Graphics for Data Analysis* (Springer-Verlag New York,  
587 2016).
- 588 49. C.-C. B. voor de Statistiek, Wijk- en buurtkaart 2019, (available at  
589 [https://www.cbs.nl/nl-nl/dossier/nederland-regionaal/geografische-data/wijk-en-](https://www.cbs.nl/nl-nl/dossier/nederland-regionaal/geografische-data/wijk-en-buurtkaart-2019)  
590 [buurtkaart-2019](https://www.cbs.nl/nl-nl/dossier/nederland-regionaal/geografische-data/wijk-en-buurtkaart-2019)).
- 591 50. P. E, B. RS, Classes and Methods for Spatial Data: the sp Package. *R News*, 9–13 (2005).
- 592 51. GitHub - rspatial/raster: R raster package, (available at  
593 <https://github.com/rspatial/raster/>).
- 594 52. cran/rgdal, (available at <https://github.com/cran/rgdal>).
- 595 53. D. S. Candido, I. M. Claro, J. G. de Jesus, W. M. Souza, F. R. R. Moreira, S. Dellicour, T. A.  
596 Mellan, L. du Plessis, R. H. M. Pereira, F. C. S. Sales, E. R. Manuli, J. Thézé, L. Almeida,  
597 M. T. Menezes, C. M. Voloch, M. J. Fumagalli, T. M. Coletti, C. A. M. da Silva, M. S.  
598 Ramundo, M. R. Amorim, H. H. Hoeltgebaum, S. Mishra, M. S. Gill, L. M. Carvalho, L. F.



- 599 Buss, C. A. Prete, J. Ashworth, H. I. Nakaya, P. S. Peixoto, O. J. Brady, S. M. Nicholls, A.  
600 Tanuri, Á. D. Rossi, C. K. V. Braga, A. L. Gerber, A. P. de C. Guimarães, N. Gaburo, C. S.  
601 Alencar, A. C. S. Ferreira, C. X. Lima, J. E. Levi, C. Granato, G. M. Ferreira, R. S. Francisco,  
602 F. Granja, M. T. Garcia, M. L. Moretti, M. W. Perroud, T. M. P. P. Castiñeiras, C. S. Lazari,  
603 S. C. Hill, A. A. de Souza Santos, C. L. Simeoni, J. Forato, A. C. Sposito, A. Z. Schreiber, M.  
604 N. N. Santos, C. Z. de Sá, R. P. Souza, L. C. Resende-Moreira, M. M. Teixeira, J. Hubner,  
605 P. A. F. Leme, R. G. Moreira, M. L. Nogueira, N. M. Ferguson, S. F. Costa, J. L. Proenca-  
606 Modena, A. T. R. Vasconcelos, S. Bhatt, P. Lemey, C.-H. Wu, A. Rambaut, N. J. Loman, R.  
607 S. Aguiar, O. G. Pybus, E. C. Sabino, N. R. Faria, Evolution and epidemic spread of SARS-  
608 CoV-2 in Brazil. *Science* (80-. ), eabd2161 (2020).
- 609 54. T. Ganyani, C. Kremer, D. Chen, A. Torneri, C. Faes, J. Wallinga, N. Hens, Estimating the  
610 generation interval for coronavirus disease (COVID-19) based on symptom onset data,  
611 March 2020. *Eurosurveillance*. **25**, 2000257 (2020).
- 612 55. B. Korber, W. M. Fischer, S. Gnanakaran, H. Yoon, J. Theiler, W. Abfalterer, N.  
613 Hengartner, E. E. Giorgi, T. Bhattacharya, B. Foley, K. M. Hastie, M. D. Parker, D. G.  
614 Partridge, C. M. Evans, T. M. Freeman, T. I. de Silva, A. Angyal, R. L. Brown, L. Carrilero,  
615 L. R. Green, D. C. Groves, K. J. Johnson, A. J. Keeley, B. B. Lindsey, P. J. Parsons, M. Raza,  
616 S. Rowland-Jones, N. Smith, R. M. Tucker, D. Wang, M. D. Wyles, C. McDanal, L. G. Perez,  
617 H. Tang, A. Moon-Walker, S. P. Whelan, C. C. LaBranche, E. O. Saphire, D. C. Montefiori,  
618 Tracking Changes in SARS-CoV-2 Spike: Evidence that D614G Increases Infectivity of the  
619 COVID-19 Virus. *Cell*. **182**, 812-827.e19 (2020).
- 620 56. E. M. Volz, *medRxiv*, in press, doi:10.1101/2020.07.31.20166082.
- 621 57. Promed Post – ProMED-mail, (available at [https://promedmail.org/promed-](https://promedmail.org/promed-post/?id=20200617.7479510)  
622 [post/?id=20200617.7479510](https://promedmail.org/promed-post/?id=20200617.7479510)).

- 623 58. Promed Post – ProMED-mail, (available at <https://promedmail.org/promed->  
624 [post/?id=7584560](https://promedmail.org/promed-post/?id=7584560)).
- 625 59. E. Cahan, COVID-19 hits U.S. mink farms after ripping through Europe. *Science* (80- ).  
626 (2020), doi:10.1126/science.abe3870.
- 627 60. (No Title), (available at <https://www.actasia.org/wp-content/uploads/2019/10/China->  
628 [Fur-Report-7.4-DIGITAL-2.pdf](https://www.actasia.org/wp-content/uploads/2019/10/China-Fur-Report-7.4-DIGITAL-2.pdf)).

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641

### 642 **Competing interests:**

643 Authors declare no competing interests.

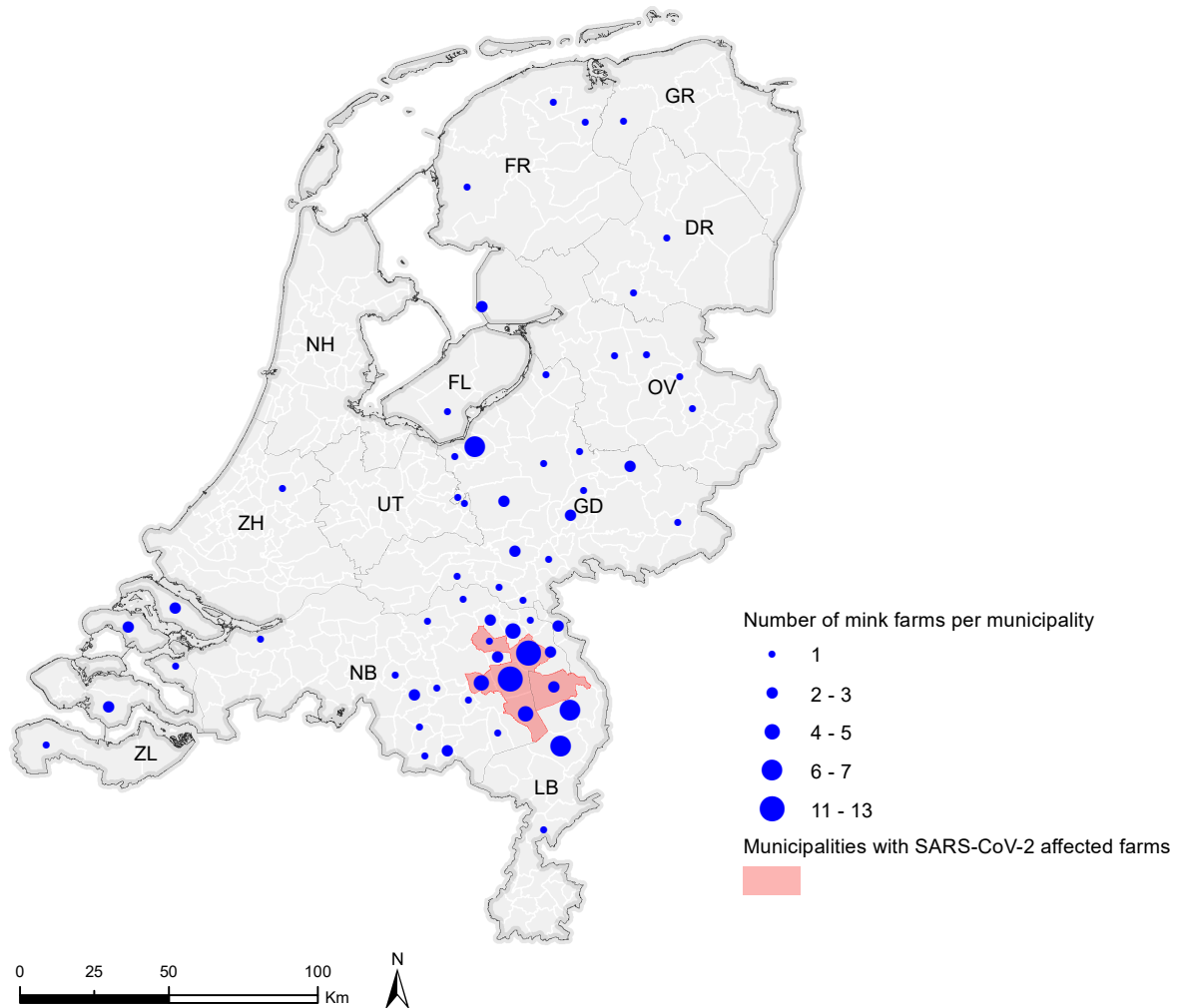
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### 645 **Data and material availability:**

646 All data, code and materials used described in this manuscript are publicly available.

647

648 **Supplements**



649

650 **Supplementary Figure 1: Number of mink farms per municipality in the Netherlands.**

651 Overview of the total number of mink farms per municipality (CBS, 2019). Municipalities

652 with SARS-CoV-2 affected farms by June 21<sup>st</sup> 2020 are shown in red.

653

654 **Supplementary table 1: GISAID acknowledgement table.**